Revision of the Malagasy lanternfly genus Belbina Stål, 1863, with two new species (Hemiptera: Fulgoromorpha: Fulgoridae)

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Abstract. The Malagasy genus Belbina Stål, 1863 (Hemiptera: Fulgoridae) is revised, transferred from the Enchophorinae Haupt, 1829 to the Aphaeninae Blanchard, 1847, and two new species, B. bourgoini sp. nov. and B. laetitiae sp. nov., are described. The genus Cornelia Stål, 1866 is proposed as a junior synonym of Belbina. The following new combinations are proposed: Belbina bergrothi (Schmidt, 1911) comb. nov. and B. nympha (Stål, 1866) comb. nov. The combination Belbina foliacea Lallemand, 1950 is restored. Aphana madagascariensis Westwood, 1851 is redescribed, transferred to Belbina and the new combination B. madagascariensis (Westwood, 1851) is proposed. Belbina vicina Lallemand, 1959 is proposed as a junior synonym of B. falleni Stål, 1863 and Cornelia atomaria (Brancsik, 1893) as a junior synonym of Belbina nympha (Stål, 1866). Neotypes are designated for B. madagascariensis (Westwood, 1851) comb. nov. and B. servillei (Spinola, 1839). The genus now comprises 12 species from Madagascar. A list of diagnostic characters, an identification key, illustrations of the male genitalia and distribution maps are provided. The falleni+ species group is defined based on characters of the male genitalia and contains the following 5 species: B. bloetei Lallemand, 1959, B. falleni Stål, 1863, B. laetitiae sp. nov., B. lambertoni Lallemand, 1922 and B. pionneau Lallemand, 1922.

Keywords. Madagascar, lanternbug, Fulgoroidea, Homoptera, planthopper

Introduction

In the process of identifying material of Fulgoridae from Madagascar, three species attributed to the genera Belbina Stål, 1863 or Cornelia Stål, 1866 were found that were not listed in the works of Lallemand (1959) or Constant (2004b). Two appear to be undescribed, while the third one represents the enigmatic Aphana madagascariensis Westwood, 1851.

The genus Belbina was described by Stål (1863a) to accommodate two species: Belbina falleni Stål, 1863 (Madagascar) and Enchophora servillei Spinola, 1839 (patria incognita, now recognized to be Madagascar). It was redescribed by Stål (1863b) later the same year, leading to misinterpretation by Metcalf (1947) who erroneously designated Enchophora sieca Walker, 1851 (South Africa) as the type species of Belbina. Lallemand (1959) validly designated Belbina falleni Stål, 1863 as the type species,
a view followed by Nast (1977) who clarified this issue according to the dates of publication of Stål’s papers.

Stål (1866) later described *Cornelia* for *C. nympha* Stål, 1866, from Madagascar.

Gerstaecker (1873) described the genus *Pyrgoteles* for *Enchophora sicca* Walker, 1851 and considered the genus close to *Belbina* Stål, 1863, and to the Oriental genera *Aphaena* Guérin-Méneville, 1834 and *Prolepta* Walker, 1851.

Karsch (1890) synonymized *Pyrgoteles* under *Belbina*. Nevertheless, in 1893 the same author described a new species in the genus *Pyrgoteles*, *P. cristatus* Karsch, 1893, thus *de facto* revalidating the genus *Pyrgoteles*.

Melichar (1908) added one species from Africa (Tanzania) to *Cornelia*: *C. usambarae*.

Schmidt (1911) described another species in *Cornelia*, *C. bergrothi* from Madagascar and placed *Cornelia* in the Aphaenini of the Fulgorinae (note that the concept of the Fulgorinae at that time equals present Fulgoridae and Aphaenini equals present Aphaeninae).

In 1918, Schumacher reviewed *Druentia* Stål, 1866 and considered *Pyrgoteles* Gerstaecker, 1873 and *Belbina* Stål, 1863 as junior synonyms of *Druentia*, the latter synonymy being invalid since *Belbina* has priority over *Druentia*. Lallemand (1922) restored *Belbina* by adding two new species from Madagascar, *B. lambertoni* (with a variety, *minuta*) and *B. pionneau*.

In his catalogue of the Fulgoridae, Metcalf (1947) listed 3 species in *Cornelia* and 4 in *Belbina* (and one variety). He included the species *sicca* Walker, 1951 in *Druentia* Stål, 1866 although he also strangely considered it the type species of *Belbina* Stål, 1863, even placing the two genera in different subfamilies: *Druentia* in the Amyclinae, Amyclini and *Belbina* in the Aphaeninae, Enchophorini.

In 1950, Lallemand added 2 species in *Belbina*: *B. recurva* and *B. foliacea* from Madagascar.

In his revision of the Afrotropical Fulgoridae, Lallemand (1959) considered *Druentia* as strictly African and *Cornelia* and *Belbina* as restricted to Madagascar. He placed the 3 genera in the subfamily Enchophorinae Haupt, 1829, which appears to be exclusively Neotropical, with *Enchophora* Spinola, 1839 as the type genus (type species, *E. recurva* (Olivier, 1891), described from Surinam; see also phylogenetic trees based on molecular data in Urban & Cryan 2009). He considered Enchophorinae a subfamily instead of following Metcalf (1947), who considered it a tribe within the Aphaeninae Blanchard, 1847. He gave a key to separate the genera based on characters such as the width of the head and the shape of the tegmina and cephalic process. He transferred *Belbina foliacea* and *Enchophora atomaria* Brancsik, 1893 to *Cornelia*, described *Belbina bloetei* and *B. vicina* (the 4 species from Madagascar), and transferred *Cornelia usambarae* Melichar, 1908 (from Tanzania) to *Druentia*. He synonymized *Belbina lambertoni* var. *minuta* Lallemand, 1922 under *B. lambertoni* Lallemand, 1922. Finally, he erroneously designated a specimen in his collection as lectotype for *Belbina servillei* (Spinola, 1839), a specimen that is not from the type series and not even the right species.

Constant (2004b) gave a key to the genera of Fulgoridae found in Madagascar (mainly based on Lallemand 1959) and a checklist of the Fulgoridae of Madagascar, including 7 species in *Belbina* and 4 species in *Cornelia*, and stated that *Cornelia* and *Druentia* could be junior synonyms of *Belbina*. He also mentioned the invalid designation by Lallemand (1959) of a lectotype for *Belbina servillei*.
This paper aims to clarify the taxonomy of the genus and the specific concept of several taxa based on illustrated morphological characters, with the description of two new species.

The current concepts of all the taxa mentioned are documented in FLOW (Bourgoin 2014).

Materials and methods

The type specimens of all species have been examined. The male genitalia were dissected as follows: the pygofer was cut from the abdomen of the softened specimen with a needle blade. It was then boiled in water for a few minutes for cleaning and to facilitate removal of pieces of tegument from the last abdominal segments. It was then dried and placed under the specimen in a gelatin capsule after examination. Endosomal characters were not used due to the difficulty to correctly inflate the membranous endosome and because it is not indispensable to separate the species in the genus Belbina. Observations were done with a Leica MZ8 stereo microscope. Pictures of specimens were taken with a Canon EOS 300 D camera with a Sigma DG Macro lens and optimized with Photoshop CS3. Pictures of genitalia were taken with a Leica DFC290 camera mounted on a Leica Z6 APO microscope, using LAS V4 software for image capture. Each series of pictures was processed with Combine ZP and optimized with Adobe Photoshop CS5.

The disc designates the central area of the tegmen, roughly covering the zone between the subcostal vein, claval joint and nodal line.

For the transcription of the labels of the types, the exact wording on each label is provided within square brackets. In the results section, species are treated in alphabetical order.

Acronyms used for the collections (name of the curator in parentheses) are as follows:

BMNH = The Natural History Museum, London, United Kingdom (M. Webb)
CAS = California Academy of Sciences, San Francisco, USA (Norman D. Penny)
FSAG = Faculté des Sciences agronomiques de Gembloux, Gembloux, Belgium (Eric Haubruege, Jeannine Bortels)
HNHM = Hungarian Natural History Museum, Budapest, Hungary (Andras Orosz)
INHS = Illinois Natural History Survey, Champaign, USA (Chris Dietrich)
MHNL = Muséum d’Histoire Naturelle de Lyon, France (Cédric Audibert)
MNHN = Muséum National d’Histoire Naturelle, Paris, France (Thierry Bourgoin, Adeline Soulier-Perkins)
MRAC = Musée royal de l’Afrique centrale, Tervuren, Belgium (Marc De Meyer, Ugo Dall’Asta)
NCSU = North Carolina State University, Raleigh, USA (Bob Blinn)
NHRS = Naturhistoriska Riksmuseet, Stockholm, Sweden (Gunvi Lindberg)
NMPC = National Museum, Praha, Czech Republic (Petr Kment)
NMW = Naturhistorisches Museum Wien, Vienna, Austria (Herbert Zettel)
RBINS = Royal Belgian Institute of Natural Sciences, Brussels, Belgium (Wouter Dekoninck)
RMNH = Nationaal Natuurhistorisch Museum (“Naturalis”), Leiden, Netherlands (Jan van Tol, Yvonne van Nierop)
SDEI = Senckenberg Deutsches Entomologisches Institut, Müncheberg, Germany (Stephan Blank)
ZIN = Russian Academy of Sciences, Zoological Institute, St. Petersburg, Russia (Vladimir Gnezdilov)
Results

Class Hexapoda Blainville, 1816
Order Hemiptera Linnaeus, 1758
Suborder Auchenorrhyncha Duméril, 1806
Infraorder Fulgoromorpha Evans, 1946
Superfamily Fulgoroidea Latreille, 1807
Family Fulgoridae Latreille, 1807
Subfamily Aphaeninae Blanchard, 1847

Morphological characters (shape of pro- and mesonotum, carinae of head, tegmina and wing venation) and especially recent molecular data (Urban & Cryan 2009) lead to the conclusion that Belbina should be transferred to Aphaeninae. The genus is close to the African genera Anecephora Karsch, 1890 and Rhicnophloea Gerstaecker, 1895 (the latter mentioned as ‘undet. Fulgoridae’ in Urban & Cryan 2009: fig. 3) and to the Oriental genera Penthicodes Blanchard, 1845 and Kalidasa Kirkaldy, 1900. The colour pattern in Belbina is very similar to that found in Penthicodes, species of Belbina looking somewhat like “Penthicodes with erected cephalic process” (see also Constant 2010 for illustrations of species of Penthicodes).

Belbina Stål, 1863


Remarks

Belbina and Cornelia are synonymized because it is not possible to find any characters allowing consistent separation of these taxa. When all species of both genera are taken into account, a gradient can be observed from the larger species (e.g., falleni and recurva) to the smaller ones (e.g., nympha and bergrothi) in all characters supposed to separate the genera, i.e., (1) the width of the vertex (supposedly narrower than eye in Cornelia and broader than eye in Belbina), (2) vertex prolonged directly on cephalic process (Cornelia) or not (Belbina) and (3) hind margin of tegmina cut transversely (Cornelia) or rounded (Belbina).

The close relationship and possible synonymy of Belbina and Druentia mentioned by Constant (2004) is not supported by the results of recent molecular studies. According to a preliminary analysis (Julie Urban pers. comm., Nov. 2013), Druentia is sister to Rhicnophloea Gerstaecker, 1895 and [Druentia + Rhicnophloea] is sister to a larger clade of Old World taxa which contains, among others, a group

Five of the twelve currently known species of Belbina seem very close and share very similar male genitalia with the following common characters: (1) pygofer, anal tube and gonostyli red or dark red; (2) gonostyli elongate with ventral margin concave in lateral view and roundly pointed apically; (3) gonostyli with strong basodorsal digitiform process directed dorsolaterally or dorsoposteriorly; (4) anal tube broadening more or less regularly from base to apex. This group is defined as the falleni+ group and contains B. bloetei, B. falleni, B. laetitiae sp. nov., B. lambertoni and B. pionneau.

**Diagnostic characters**

The genus can be recognized by the following combination of characters: (1) head much narrower than pronotum; (2) frons longer than broad, with longitudinal carinae; (3) cephalic process present, about as long as frons and projecting anterodorsally to posterodorsally; (4) lateral carinae of cephalic process (i.e., prolongation of the lateral carinae of frons and of vertex) fused in a single longitudinal carina before apex of process; (5) pronotum with a strong tectiform median carina and a deeply impressed point on each side of it.

*Belbina* is very similar to *Druentia* but can be separated by character (4): in *Druentia* the lateral carinae of the vertex reach the apex of the cephalic process without fusing together.

**Sexual dimorphism**

Males 15 to 30% smaller in size than females.

**Distribution**

Only recorded from Madagascar.

**Identification key to the species of Belbina**

The disc of the hind wings can be either orange or red in some species (observed in *B. bergrothi*, *B. madagascariensis* and *B. nympha*). It is possible that other species presently known only from red hind-winged specimens also have ones with orange hind wings.

The red colour of the abdomen can also fade to orange or yellow in collection specimens, especially if they have been preserved in ethanol.

1. Hind wings red or orange on disc (Figs 1A, 2A) .................................................................2
   – Hind wings with disc bluish, greenish or brown (Figs 3A, 6A) ............................................10

2. Ground colour of tegmina orange or red (Figs 1A, 7A, 10A) .................................................3
   – Ground colour of tegmina brown to pale rosy brown (Figs 2A, 4A) .................................5

3. Mesonotum with 2 black patches on disc (Fig. 1C); small-sized (less than 22 mm long) ........
   .........................................................................................................................*Belbina bergrothi* (Schmidt, 1911)
   – Mesonotum without black patches (Figs 7C, 10C); larger (more than 24 mm long) ..............4

4. Tegmina orange to red with minute black spots; 3 large black patches on costal cell and one at
   the base of clavus (Fig. 7A) .........................................................................................*Belbina lambertoni* Lallemand, 1922
6. Tegmina dark red with membrane pale brown; numerous, irregular, blackish brown spots (Fig. 10A) .................................................................................................................. Belbina pionneai Lallemand, 1922

5. Head with a foliaceous process above the eye, higher than diameter of eye in side view; median carina of pronotum strongly sinuate and elevated posteriorly in side view (Fig. 5D) .... Belbina foliacea Lallemand, 1959

   - Head without large foliaceous process above the eye, process reduced to a lamina not higher than half of diameter of eye; median carina of pronotum not strongly sinuate and elevated posteriorly in side view (Figs 4D, 11D) ........................................6

6. Small-sized, less than 22 mm long; cephalic process directed anterodorsad (Fig. 9D); abdomen with 2 dorsal rows of black spots; hind wings with 4–6 little black spots on red zone (Fig. 9A) .................................................................................. Belbina nympha (Stål, 1866)

   - Larger, more than 26 mm long; cephalic process angulously directed dorsad or posteriorad (Fig. 2D), or, if not, dorsum of abdomen red without rows of black spots; hind wings with 2 small black spots or larger black spots (Figs 4A, 8A) ...................................................7

7. Cephalic process narrow, elongate, obliquely directed anterodorsad (Fig. 8D); blackish brown spots with pale center on tegmina (Fig. 8A) ........ Belbina madagascariensis (Westwood, 1851)

   - Cephalic process broad, strongly curved dorsad or posteriorad (Fig. 2D); dark spots on tegmina without pale center (Figs 2A, 4A) ...................................................................................................8

8. Cephalic process projecting dorsad, apex not surpassing level of anterior margin of eye in lateral view (Fig. 2D); abdomen without rows of black spots dorsally; no large black patch on basal half of claval joint; no paler transverse patch at half length of tegmen (Figs 2A, 4A) ...9

   - Cephalic process more strongly curved and projecting posterodorsad, reaching level of posterior margin of eye in lateral view (Fig. 11D); abdomen with 2 rows of blackish spots dorsally; one large black elongate patch on basal half of claval joint; one transverse paler patch at about half of length of tegmen (Fig. 11A) ................. Belbina recurva Lallemand, 1950

9. Ground colour of tegmina and head pale rosy brown; a rectangular paler patch on clavus (Fig. 2A) ................................................................................................................. Belbina bloetel Lallemand, 1959

   - Ground colour of tegmina and head brown; no evident paler patch on clavus (Fig. 4A) .............. Belbina falleni Stål, 1863

10. Head, pro- and mesonotum red (Fig. 12C–E); ground colour of tegmina bluish green (Fig. 12A) .................................................................................................................. Belbina servillei (Spinola, 1839)

   - Head, pro- and mesonotum brown (Figs 3C–D; 6C–D); ground colour of tegmina bluish green or brownish (Figs 3A, 6A) .......................................................... Belbina laetitia sp. nov.

11. Ground colour of tegmina bluish green; hind wings nearly totally infuscate with a suturo-basal, not well limited zone, and spots, paler, milky (Fig. 6A); larger (24 mm long in males, 27 mm long in females) .............................................................................. Belbina bourgoini sp. nov.

   - Ground colour of tegmina brownish with a dark reddish band after half of length; hind wings with paler, bluish green disc very distinctly separated from blackish apex and sutural margin (Fig. 3A); smaller (19 mm long in males) .............................................. Belbina bourgoini sp. nov.
Belbina bergrothi (Schmidt, 1911) comb. nov.
Figs 1A–E, 13–14, 36, 46

Cornelia bergrothi Schmidt, 1911: 242 (type in SDEI).


**Diagnostic characters**

(1) disc of hind wings orange or red (Fig. 1A); (2) 2 black patches on mesonotum (Fig. 1C); (3) size less than 22 mm long; (4) cephalic process directed dorsal, narrow in anterior view (Fig. 1D–E).

LT: ♂ (n = 3) 17.8 mm (17.0–19.0); ♀ (n = 2) 20.5 mm (20.0–21.0).

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Material examined

Holotype
MADAGASCAR: ♀ (on photographs), [Madagasc. Amber Geb.] [Holotypus] [Cornelia Bergrothi Schmidt ♀ Edm. Schmidt determ. 1911] [Coll. Breddin] [Dtsch. Entomol. Institut Berlin], Montagne d’Ambre (Ambositra), 12°30’ S, 49°10’ E (SDEI).

Additional material
MADAGASCAR: 1 ♂, 1 ♀, E Madagascar, Massif du Marozejy (rés. nat. intégr. 12), 1300 m, 2–8 Dec. 1972, A. Peyrieras, 14°26’ S, 49°44’ E (MNHN); 1 ♂, same data, 4 Dec. 1972 (RBINS); 1 ♀, same data, 12 Dec. 1972 (RBINS); 1 ♀, Fianarantsoa province, Parc National Ranomafana, Belle Vue et Talatokely,

21°15.99’ S 47°25.21’ E, 1020 m, 21–28 Jan. 2002, Malaise trap, secondary forest, R. Harin’Hala (CAS); 1 ♂, Moramanga region, Andaside, PL hôtel, 18°56.820’ S, 48°25.155’ E, 22 Nov. 2005, 930 m, Bourgojn, Ouvrard, Attié, Soulíer-Perkins (MNHN); 1 ♀, no data (FSAG); 1 ♀, Antananarivo province, 46 km NE of Ankazobe, Ambohitantely, 18°11.88’ S, 47°16.89’ E, Malaise trap in sclerophyl forest, 700 m, 16–27 Jan. 2005, Irvin & Harin’Hala (CAS); 1 ♂, route d’Anosibe, 11 Nov. 1961 (MHNL).

Examined on photograph
MADAGASCAR: 1 specimen, Vohimana, 4 Jan. 2006, 13:35, Nicolas Cliquennois (Fig. 36), 18°55’ S, 48°30’ E.

Male genitalia
Black with gonostyli showing large, basoventral ochraceous patch (Fig. 13); pygofer higher than long and with posterior margin sinuate in lateral view (Fig. 13); anal tube elongate, 1.6 times longer than broad at apex and with lateral margins sinuate in dorsal view (Fig. 14); gonostyli elongate, 2.4 times longer than high, not surpassing apex of anal tube and acutely rounded at apex in lateral view (Fig. 13); dorsal margin of gonostyli projecting laterally from base to midlength and with strong tooth directed ventrally (Figs 13–14); ventral margin and apical half of dorsal margin broadly rounded in lateral view (Fig. 13).

Remarks
Male genitalia without basodorsal process on gonostyli. *Belbina bergrothi* can be separated (1) from *B. recurva* by not having the margins of the anal tube produced into a semi-circular lateral plate; (2) from *B. madagascariensis* and *B. nympha* by having the gonostyli acutely rounded apically and more elongate; (3) from *B. foliacea* by not having the gonostyli angulose above mediadorsal tooth.

Distribution
See Fig. 46.

**Belbina bloetei** Lallemand, 1959
Figs 2A–E, 15–16, 46

*Bellina bloetei* Lallemand, 1959: 92, fig. 40a–b (lateral view of head, wing venation) (type in RMNH).

*Belbina bloetei* – Constant 2004b: 31 (listed).

Note
The species epithet refers to the Dr H.C. Blöte (RMNH) and is emended following the article 32.5.2.1 of ICZN (1999).

Diagnostic characters
(1) disc of hind wings red (Fig. 2A); (2) size more than 26 mm; (3) cephalic process projecting dorsad, broad (Fig. 2D–E); (4) ground colour of tegmina and head pale rosy brown; (5) rectangular pale patch on clavus (Fig. 2A, C).

LT: ♂ (n = 1) 27.0 mm; ♀ (n = 1) 29.5 mm.
CONSTANT J., Fulgoridae of Madagascar: \textit{Belbina} (Hemiptera)

Material examined

Holotype
MADAGASCAR: ♀ (examined on photographs), [Madagascar, coll. Lambert.] [environ de Tananarive] [Type] \textit{Belbina blotei} Lall., V. Lallemand det., 195 [RMNH.INS.780532], Tananarive (Antananarivo), 18°55’ S, 47°31’ E (RMNH). – Note: “Lambert” = Lamberton.

Paratypes
MADAGASCAR: 1 ♀ (examined on photographs), [Madagascar, rec. Lamberton] [environ de Tananarive] \textit{Belbina blotei} Lallemand, V. Lallemand det., 195 [Paratype] [RMNH.INS.780531] (RMNH); 1 ♂, 1 ♀, [environ de Tananarive] (FSAG). Note: Lallemand (1959) stated that all types are deposited in RMNH but he actually retained 2 paratypes in his collection (now in FSAG).

Additional material
MADAGASCAR: 1 ♂, Mahajanga Prov., Forêt de Tsimembo, 8.7 km 336° NNW of Soatana, 19°1’17” S, 44°26’26” E, 20 m, 21–25 Nov. 2001, at light in tropical dry forest, Fisher, Griswold \textit{et al.} (CAS); 2 ♀♀, near Tananarive, Lambert (RBINS); 2 ♀♀, Morondava à Mahabo, Last 1854-91, 20°20’ S, 44°28’30” E (MNHN); 1 ♂, Madagascar (NMPC); 1 ♂, no data (NCSU).

Male genitalia
Dark red (Figs 15–16); pygofer higher than long and with posterior margin concave in middle in lateral view (Fig. 15); anal tube slightly elongate, 1.16 times longer than broad at apex and with lateral margins bisinuate in dorsal view (Fig. 16); margin of anal opening pointed posteriorly (Fig. 15) and hiding lateral margin apically in dorsal view (Fig. 16); gonostyli elongate, 1.18 times longer than high (dorsal process included), strongly surpassing apex of anal tube and acutely rounded at apex in lateral view (Fig. 15); ventral margin straight on basal $\frac{1}{5}$, then strongly excavate (Fig. 15); dorsal margin with basal strong digitiform process projecting laterodorsally, strong hook laterally at middle of process projecting posterovertrally and posterior margin of process excavate between apex and hook (Fig. 15); apex of digitiform process rounded and broad in dorsal view (Fig. 16).

Remarks
\textit{Belbina blotei} is a member of the \textit{falleni+} group, showing strong basodorsal digitiform process on gonostyli. It can be separated (1) from \textit{B. falleni}, \textit{B. laetitiae} sp. nov., \textit{B. lambertoni} and \textit{B. pionneaui} by the more strongly concave ventral margin of the gonostyli in lateral view; (2) from \textit{B. falleni} by the less excavate dorsal margin of the gonostyli in dorsal view; (3) from \textit{B. laetitiae} sp. nov. and \textit{B. lambertoni} by the more acute apex of the gonostyli in lateral view; (4) from \textit{B. pionneaui} by the more rounded apex of the basodorsal digitiform process of the gonostyli.

Distribution
See Fig. 46.

\textit{Belbina bourgoini} sp. nov.
Figs 3A–E, 17–18, 47
\url{urn:lsid:zoobank.org:act:266C852F-6F78-48F8-AC72-3A286E350836}

Diagnostic characters
(1) disc of hind wings light white-bluish (Fig. 3A); (2) small-sized (less than 22 mm long); (3) tegmina brown with dark red-brown band (Fig. 3A); (4) head, pro- and mesonotum brown (Fig. 3C–E).
Etymology

The species is dedicated to Prof. Thierry Bourgoin (MNHN), in acknowledgement of his permanent help and support from the beginning of the work of the author.

Material examined

Holotype


Paratypes


Description

LT: ♂ (n = 2) 19.2 mm (19.0–19.5).

Head. Yellow-brown sometimes suffused with red (Fig. 3C–E); vertex with hind margin and sides strongly carinate (Fig. 3C); sides of vertex bisinuate in lateral view (Fig. 3D); frons with 2 carinae extending on sides of cephalic process (Fig. 3E); cephalic process about 1.5 times as long as diameter of eye, projecting dorsad to posterodorsad (Fig. 3D); apical half carinate anteriorly and strongly emarginate laterally (Fig. 3E); posterior side of process with 2 longitudinal, sinuate carinae; lateral oblique carina between vertex and frons before eye (Fig. 3D); postclypeus with sides strongly carinate and obsolete median carina; anteclypeus with strong median carina (Fig. 3E); labium very elongate, surpassing hind coxae but not reaching apex of abdomen.

Thorax. Yellow-brown varied with irregular paler spots (Fig. 3C–D); pronotum carinate anteriorly; sinuate carinae at anterior margin of disc, reaching anterior margin before median carina (Fig. 3C); median carina strong, sinuate in lateral view; hind margin strongly elevated above level of mesonotum medially (Fig. 3D); impressed point on disc on each side of carina; obsolete tubercles at each side of disc (Fig. 3C); sides of prothorax longitudinally carinate; lateral lobes of pronotum rounded posteriorly (Fig. 3D–E); mesonotum with median carina stopped at scutellum posteriorly; strongly sinuate carina on each side of disc, Y-shaped anteriorly; scutellum transversely wrinkled (Fig. 3C).

Tegmina (Fig. 3A). Pale yellow-brown with irregular brown-black markings; ground colour red-brown on middle; clavus paler; white marking along sutural margin at level of nodal line; costal margin slightly rounded; apical angles rounded; sutural and apical margins sinuate, the latter oblique; ratio LTg/BTg = 2.6.

Hindwings (Fig. 3A). Black-brown with large, light white-buish discal patch of basal half, reaching costal margin and extending posteriorly along vein Cu without reaching sutural margin; black-brown elongate marking basally along vein M; big black spot along vein PCu followed by 1–2 smaller spots; apex rounded, sutural margin sinuate.

Legs (Fig. 3A). Elongate and slender; femora brown with 3–4 narrow, sinuate and often incomplete pale yellow rings; pro- and mesotibiae brown with 2 pale yellow rings; metatibiae brown with pale yellow markings at spines; pro- and mesotarsi black-brown; first metatarsomere pale yellow-brown on basal ⅔; rest of metatarsi brown; metatibiae with 6–7 lateral and 7 apical spines; first metatarsomere with 9 apicoventral spines.
CONSTANT J., Fulgoridae of Madagascar: *Belbina* (Hemiptera)

**Abdomen** (Fig. 3A). Brown, darker ventrally.

**Male genitalia**

Dark brown (Figs 17–18); pygofer higher than long and with posterior margin nearly straight, slightly sinuate dorsally in lateral view (Fig. 17); anal tube elongate, 1.6 times longer than broad at apex and with lateral margins bisinuate in dorsal view (Fig. 18), slightly curved ventrally and with apex broadly rounded in lateral view (Fig. 17); gonostyli elongate, 1.6 times longer than high (dorsal process included), surpassing apex of anal tube and broadly rounded at apex in lateral view (Fig. 17); ventral margin straight, with strong angle at basal \( \frac{1}{8} \) (Fig. 17); dorsal margin with basal slender digitiform process projecting laterodorsally, pointed tooth laterally at middle of process projecting anteroventrally, apex of digitiform process narrowing apically (Fig. 17); apical \( \frac{3}{5} \) of dorsal margin sinuate (Fig. 17).

**Remarks**

*Belbina bourgoini* sp. nov. can be separated (1) from all species of the *B. falleni*+ group by the brown colour of the genitalia, the more slender basodorsal digitiform process on the gonostyli and by having the gonostyli rounded at the apex, with the dorsal margin sinuate after the process; (2) from *B. bergrothi*, *B. foliacea*, *B. madagascariensis* and *B. nympha* by the basodorsal digitiform process on the gonostyli; (3) from *B. servillei* by having the digitiform basodorsal process of the gonostyli slender and narrow, not laminate.

**Distribution**

See Fig. 47.

*Belbina falleni* Stål, 1863

Figs 4A–E, 19–20, 37–38, 47

*Belbina falleni* Stål, 1863a: 233 (type in NMW).

*Belbina vicina* Lallemand, 1959: 90, fig. 38 (key, description and dorsal view of head and thorax) (type in FSAG). syn. nov.


*Belbina vicina* – Constant 2004b: 31 (listed).

**Diagnostic characters**

(1) disc of hind wings red with black markings (Fig. 4A); (2) ground colour of tegmina pale brown (Fig. 4A); (3) sides of vertex laminate above eye but not foliaceous (Fig. 4C–D); (4) large-sized (more than 26 mm long); (5) cephalic process broad, strongly curved dorsad (Fig. 4D); (6) clavus with colour pattern similar to the rest of the tegmen (Fig. 4A).

LT: ♂ (n = 5) 26.6 mm (26.0–27.0); ♀ (n = 7) 29.1 mm (28.5–30.2).

**Material examined**

**Type material**

MADAGASCAR: Holotype of *Belbina falleni*, ♂, [Madagascar, Coll. Signoret] [Falléni det. Signoret] [Type] (NMW).

MADAGASCAR: Holotype of *Belbina vicina* Lallemand, 1959, ♂, [Manjakandriana, Madagascar] [Type.] [*Belbina vicina* Lall., V. Lallemand det., 1957], 18°55’ S, 47°48’ E (FSAG).
Additional material

MADAGASCAR: 1 ♀, Manjakandriana (FSAG); 2 ♂♂, 1 ♀, near Tananarive, Lambert, Tananarive (Antananarivo), 18°55' S, 47°31' E (RBINS); 2 ♂♂, Vohemar (Iharaoa), 13°21’ S, 50°00’ E (RBINS, NCSU); 1 ♂, N Madagascar (BMNH); 1 ♀, Montagne d’Ambre, Feb. 1930, Sicard, Montagne d’Ambre (Ambohitra), 12°30’ S, 49°10’ E (MNHN); 1 ♀, no locality, Dec. 1930 (MNHN); 1 ♀, no data (NCSU).

Examined on photographs

MADAGASCAR: 1 specimen, Montagne d’Ambre, 27 Oct. 2009, 15:05, Nicolas Cliquennois (Fig. 38); 1 specimen, Montagne d’Ambre, Joffreville (Ambohitra), 2009, Rhett A. Butler (Fig. 37).

Male genitalia

Dark red (Figs 19–20); pygofer higher than long and with posterior margin sinuate in lateral view (Fig. 19); anal tube slightly elongate, 1.27 times longer than broad at apex and with lateral margins bisinuate in dorsal view on apical half (Fig. 20); posterior margin notched in lateral view (Fig. 19), with hind margin of anal opening projecting posteriorly (Fig. 19) and hiding lateral margin apically in dorsal view (Fig. 20); gonostyli elongate, 1.36 times longer than high (dorsal process included), strongly surpassing anal tube and acutely rounded at apex in lateral view (Fig. 19); ventral margin straight on basal 1/5, then slightly sinuate (Fig. 19); dorsal margin with basal strong digitiform process projecting laterodorsally, strong hook laterally at middle of process projecting posteroventrally and posterior margin of process excavate between apex and hook (Fig. 19); apex of digitiform process rounded and broad in lateral view (Fig. 19), slightly compressed laterally in dorsal view (Fig. 20).

Remarks

Lallemand (1959) erroneously mentioned that the type of B. vicina is a female. However it is obvious that the type is a male because he stated that “genitalia have similar shape as those of B. pionneaui and B. lambertoni”.

Lallemand (1959) described B. vicina based on a specimen of B. falleni presenting the cephalic process more strongly curved posteriorad than in another specimen he had identified in his collection as B. falleni; both specimens were from the same locality (Manjakandriana). Male genitalia are similar and intraspecific variability in the direction of the cephalic process has been observed from series in several species of Belbina. For those reasons, B. vicina is here synonymized with B. falleni.

Belbina falleni is a member of the falleni+ group, showing a strong basodorsal digitiform process on the gonostyli. It can be separated (1) from B. bloetei by the less concave ventral margin of the gonostyli in lateral view; (2) from B. laetitiae sp. nov. and B. lambertoni by the laterally more strongly compressed digitiform process and the more acutely rounded apex of the gonostyli in lateral view; (3) from B. pioneae by the more rounded apex of the basodorsal digitiform process of the gonostyli and the more strongly notched ventroapical margin of the anal tube under the anal opening in lateral view.

Distribution

See Fig. 47.
**Belbina foliacea** Lallemand, 1950 comb. rev.
Figs 5A–E, 21, 47

*Belbina foliacea* Lallemand, 1950: 84 (type in FSAG).

*Cornelia foliacea* – Lallemand 1959: 87, fig. 32a–c (key, description, lateral view of head and male genitalia). — Constant 2004b: 31 (listed).

**Diagnostic characters**
1. disc of hind wings red (Fig. 5A); 2. carina above eye strongly expanded dorsad into foliaceous process (Fig. 5C–E); 3. cephalic process strongly curved dorsad (Fig. 5D); 4. tegmina variegated brown-olivaceous; 5. veins of tegmina elevated (Fig. 5A).

**Material examined**

**Holotype**
MADAGASCAR: ♂, [Maromandia, Madagascar, Lamberton] [Type.] *Cornelia foliacea* Lall., V. Lallemand det., 1954], 18°56’S, 49°03’ E (FSAG).

**Paratype**
MADAGASCAR: 1 ♀, no label, probably same data as holotype.

**Male genitalia**
The male genitalia were dissected and drawn by Henry Synave for Lallemand’s revision (1959). Only one gonostylus and the phallic complex were illustrated. The organs were preserved in a small glass vial under the specimen. The anal tube is missing and the pygofer is severely damaged. As it has been treated with potassium hydroxide, it is not possible to know the original colour.

Gonostyli elongate, 1.7 times longer than high and broadly rounded at apex in lateral view; ventral margin nearly straight; dorsal margin strongly sinuate basally and with a strong angle at basal third, then straight; strong lateral tooth directed posteroventrally along dorsal margin under angle (Fig. 21).

**Remarks**
Male genitalia without a baso-dorsal process on the gonostyli.

*Belbina foliacea* can be separated from *B. bergrothi*, *B. madagascariensis*, *B. nympha* and *B. recurva* by the strong angle above the medio-dorsal tooth and the broadly rounded apex of the gonostyli.

**Distribution**
See Fig. 47.
Diagnostic characters
(1) disc of hind wings milky, base and anal lobe suffused with red (Fig. 6A); (2) head, pro- and mesonotum brown (Fig. 6C–E); (3) ground colour of tegmina bluish green (Fig. 6A); (4) large-sized (more than 24 mm long); (5) cephalic process broad, projecting dorsad to posterodorsad (Figs 6D–E).

LT: ♂ (n = 8) 24.3 mm (23.2–25.4); ♀ (n = 11) 26.8 mm (25.1–28.4).

Etymology
Dedicated to Mrs Laetitia Despontin, the mother of my children Emilie and Guillaume.

Material examined

Holotype

Paratypes
Examined on photograph
MADAGASCAR: 1 specimen (Figs 43–45), Ile Sainte Marie, 25 Nov. 2011, Paul Bertner, 16°50’ S, 49°55’ E.

Description

HEAD. Brown, sometimes slightly suffused with red (Fig. 6C–E); vertex with hind margin strongly carinate and sides laminate, bisinuate in lateral view and elevated above eye (Fig. 6C–D); frons longitudinally wrinkled with 2 carinae extending on sides of cephalic process (Fig. 6E); cephalic process slightly more than twice as long as diameter of eye, projecting posterodorsad to dorsad (Fig. 6D); apical half of process lanceolate in anterodorsal view and with median carina anteriorly (Fig. 6C, E); posterior side of process with 2 carinae fused before apex; strongly sinuate carina under side of vertex, not reaching eye (Fig. 6D); postclypeus with sides strongly carinate and median carina; anteclypeus with strong median carina (Fig. 6E); labium very elongate, surpassing hind coxae but not reaching apex of abdomen.

THORAX. Brown with mesonotum and meso- and metapleura red (Fig. 6C–E); pronotum with strong median carina sinuate in lateral view (Fig. 6C–D); hind margin strongly elevated above level of mesonotum medially; sides of prothorax longitudinally carinate; lateral lobes of prothorax rounded posteriorly (Fig. 6D); mesonotum (Fig. 6C) with median carina stopped posteriorly before scutellum; strongly sinuate carina on each side of disc, sometimes divided into external, sinuate carina and internal, angulous carina; scutellum transversely wrinkled.

TEGMINA (Fig. 6A). Green-brown to pale olivaceous-brown with irregular, dense, black-brown to black markings; apical ⅓ brown-black with large, unprecise, darker markings and some minute, whitish spots; small white patch along sutural margin at nodal line with black spot in middle; costal and apical margins slightly rounded, the latter oblique; apical angles rounded; sutural margin sinuate.

HIND WINGS (Fig. 6A). Smoky brown-black, darker on apical ⅃; anal lobe and base suffused with red; milky patch on basal ⅔ from costal margin to vein M or Cu; veins darker than ground colour; 4–6 dark brown-black ocelli on clavus with round, waxy spot in middle; irregular white waxy spots on apical ⅔ and on disc; apex rounded, sutural margin emarginate at ⅔.

Figs 43–45. Belbina laetitiae sp. nov. in natura, Ile Sainte Marie, 25 Nov. 2011, photographs by Paul Bertner.
Legs (Fig. 6A). Elongate and slender; dark brown-black, with obsolete pale yellow-brown rings often only marked by small spots: 3–4 rings on pro- and mesofemora, 2 on mesotibiae and on metafemora; spines of metafemora with pale yellow-brown spots near base; metatibiae with 5 lateral and 7 apical spines; 12 spines apicoventrally on first metatarsomere.

Abdomen (Fig. 6A). Bright red with 2 longitudinal rows of 4 black spots dorsally.

Male genitalia

Red (Figs 22–23); pygofer higher than long and with posterior margin slightly sinuate in lateral view (Fig. 22); anal tube slightly elongate, 1.39 times longer than broad at apex and with lateral margins nearly straight in dorsal view (Fig. 23); posterior margin strongly notched in lateral view (Fig. 22), with hind margin of anal opening projecting posteriorly and pointed in lateral view (Fig. 22), and hiding lateral margin apically in dorsal view (Fig. 23); gonostyli elongate, 1.22 times longer than high (dorsal process included, 1.72 times longer than high without process), surpassing anal tube and acutely rounded at apex in lateral view (Fig. 22); ventral margin straight on basal \( \frac{1}{5} \), then slightly sinuate (Fig. 22); dorsal margin, with basal strong digitiform process projecting laterodorsally and slightly curved posteriorly, strong hook laterally at middle of process projecting anteroventrally (Fig. 22); apex of digitiform process rounded and broad (Fig. 22–23).

Remarks

Belbina laetitiae sp. nov. is a member of the falleni+ group, showing a strong basodorsal digitiform process on the gonostyli. It can be separated (1) from B. bloetei by the less concave ventral margin and more rounded apex of the gonostyli in lateral view; (2) from B. falleni by the round cross section of the digitiform process and the more rounded apex of the gonostyli in lateral view; (3) from B. lambertoni by having an anal tube that is more strongly curved posteriorly in lateral view and more elongate in dorsal view; (4) from B. pionneau by the more rounded apex of the basodorsal digitiform process of the gonostyli and the more strongly notched ventroapical margin of the anal tube under the anal opening in lateral view.

Distribution

See Fig. 48.

Belbina lambertoni Lallemand, 1922

Figs 7A–E, 24–25, 39, 49

Belbina lambertoni Lallemand, 1922: 62 (type in FSAG).
Belbina lambertoni var. minuta Lallemand, 1922: 62 (type in FSAG).

Belbina lambertoni var. minuta – Lallemand 1959: 91 (synonymised with B. lambertoni Lallemand, 1922).

Diagnostic characters

(1) disc of hind wings red (Fig. 7A); (2) tegmina orange to red with 3 large black patches on costal cell and one at base of clavus (Fig. 7A); (3) cephalic process strongly curved dorsad (Fig. 7D); (4) large-sized (more than 24 mm long); (5) head orange (Fig. 7C–E).

LT: ♂ (n = 12) 27.8 mm (24.6–30.1); ♀ (n = 17) 34.5 mm (32.5–38.2).
Material examined

Holotype
MADAGASCAR: ♀, [Manjakandriana, Madagascar] [Type] [Belbina lambertoni] Lall, V. Lallemand det., 195] (FSAG).

Paratypes
MADAGASCAR: 5 ♂♂, 2 ♀♀, [Manjakandriana, Madagascar] [Paratype] (FSAG; 1 ♀: MRAC); 1 ♀, [Manjakandriana, Madagascar] [Type] [Belbina lambertoni] Lall, V. Lallemand det., 1959] (NHRS-HMEM 000000104) (NHRS); 1 ♀, [Manjakandriana, Madagascar] [Paratype] [Musée du Congo] [Belbina lambertoni] Lall, V. Lallemand det., 195], 18°55’ S, 47°48’ E (MRAC).

Note: Lallemand (1922, 1959) stated that the type and paratypes are in his collection. It seems, however, that he subsequently gave one paratype to NHRS and two to MRAC.

Additional material
MADAGASCAR: 2 ♂♂, 2 ♀♀, near Tananarive, Lamberton, Tananarive (Antananarivo), 18°55’ S, 47°31’ E (FSAG); 1 ♂, 5 ♀♀, Vohemar, coll. Le Moul, Vohemar (Ihararaoa), 13°21’S, 50°00’ E (FSAG, 4 ♀♀: NCSU); 2 ♂♂, Maroantsetra Prov., Parc National Tsingy de Bemaraha, 2.5 km 62° ENE of Bekopaka, Ankidrodroa River, 19°75’6” S, 44°48’53” E, 100 m, 11–15 Nov. 2001, tropical dry forest on Tsingy, Fisher, Griswold et al. (CAS); 1 ♂, 10 ♀♀, Tananarive (5 ♀♀: NCSU; 1 ♂, 4 ♀♀: RBINS; 1 ♀: MNHN); 5 ♂♂, 3 ♀♀, Morondava à Mahabo, Last 1854-91, 20°20’ S, 44°28’30” E (MNHN); 1 ♀, Antalaha region, Dec. 1935, Vadon, 14°53’ S, 50°17’ E (MNHN); 3 ♀♀, Madagascar, coll. De Bergevin (MNHN); 1 ♂, Vohemar, coll. de Bergevin (MNHN); 1 ♀, Ambatofitorano, Jul. 1978, 20°49’ S, 47°11’ E (RBINS); 2 ♂♂, Ankazoabo, SW Madagascar, K.U. Tyduna, 22°17’ S, 44°31’ E (ZIN); 1 ♂, Madagascar (NMW); 1 ♀, Maroantsetra, coll. Le Moul, 15°26’ S, 49°44’ E (NMW); 1 ♂, Madagascar, 20 Oct. 1905, Dr. Kiderlen (NMW); 1 ♂, 2 ♀♀, no data (NCSU); 2 ♂♂, 4 ♀♀, Antsalova, Dec. 1992, 18°24’ S, 44°22’ E (MHNL); 2 ♂♂, Madagascar (MHNL); 2 ♀♀, idem (INHS); 1 ♀, Miandrivasa (Miantirvazo), Mar. 1961, 19°33’20” S, 45°27’03” E (MHNL); 1 ♂, 3 ♀♀, no label (NCSU).

Examined on photograph
MADAGASCAR: 1 ♂, Berenty Reserve, riverine forest, 25°00’20” S, 46°18’10” E, 15 Nov. 2008, Jason Cryan (Fig. 39).

Male genitalia
Bright red (Figs 24–25); pygofer higher than long and with posterior margin bisinuate in lateral view (Fig. 24); anal tube slightly elongate, 1.1 times longer than broad at apex and with lateral margins slightly bisinuate, slightly concave on apical half in dorsal view (Fig. 25); posterior margin slightly notched in lateral view (Fig. 24), with hind margin of anal opening pointed posteriorly in lateral view (Fig. 24) and hiding lateral margin apically in dorsal view (Fig. 25); gonostyli elongate, 1.12 times longer than high (dorsal process included), surpassing anal tube and rounded at apex in lateral view (Fig. 24); ventral margin straight on basal ¼, then slightly sinuate (Fig. 24); dorsal margin with basal, strong digitiform process projecting laterodorsally, strong hook laterally at middle of process projecting posteroventrally and posterior margin of process sinuate between apex and hook (Fig. 24); apex of digitiform process rounded and broad in lateral and dorsal view (Figs 24–25).

Remarks
Belbina lambertoni is a member of the falleni+ group, showing a strong basodorsal digitiform process on the gonostyli. It can be separated (1) from B. bloetei by the less concave ventral margin and more rounded apex of the gonostyli in lateral view; (2) from B. falleni by the round cross section of the digitiform process and the more rounded apex of the gonostyli in lateral view; (3) from B. laetitiae sp. nov. by the anal tube being less curved posteriorly in lateral view and less elongate in dorsal view; (4)
from *B. pionneauti* by the more rounded apex of the basodorsal digitiform process of the gonostyli and the more strongly notched ventroapical margin of the anal tube under the anal opening in lateral view.

**Distribution**

See Fig. 49.

*Belbina madagascariensis* (Westwood, 1851) comb. nov.
Figs 8A–E, 26–27, 46

*Aphana madagascariensis* Westwood, 1851: 208 (neotype in MNHN).


**Diagnostic characters**

(1) disc of tegmina red or orange (Fig. 8A); (2) cephalic process elongate and narrow, projecting anterodorsad (Fig. 8D–E); (3) ground colour of tegmina brown (Fig. 8A); (4) large-sized (more than 26 mm long); (5) black-brown spots with white center on tegmina (Fig. 8A).

**Material examined**

*Neotype*


Note: A neotype is here designated in order to stabilise the nomenclature in the group, following rule 75.3 of the International Code of Zoological Nomenclature (ICZN 1999). The specimen chosen here is probably the one on which Westwood (1851) based his description. It was collected by Jules Prosper Goudot in 1832 and included in the collections of the MNHN in 1834 (Adeline Soulier-Perkins pers. comm.). Westwood (1851) stated that the specimen he examined was from the collections of the “Mus. Jardin des Plantes, Paris”, the name of the MNHN at that time. No specimen labelled *Aphana madagascariensis* was found in the collections of the MNHN, but the specimen chosen here perfectly matches the original description. Lallemand (1959) assumed that the type was lost and stated that, according to the description, the species probably belonged to *Belbina* or *Cornelia*. As it is not totally certain that the specimen is the one on which the original description was based, I designate it here as a neotype rather than recognize it as the holotype.

*Additional material*

MADAGASCAR: 1 ♀, Mad[agascar] (MNHN); 1 ♀, Tenina, J. Vadon, 15°45’ S, 49°40’ E (RBINS); 1 ♂, NE Madagascar, Fampanambo, Jul. 1959, J. Vadon, 15°22’ S, 49°38’ E (MRAC).

**Redescription**

LT: ♂ (n = 1) 26.3 mm; ♀ (n = 1) 28.3 mm.

**Head.** Yellow-brown with 2 darker patches on disc of vertex (Fig. 8C–E); yellow-orange patch around insertion of antennae (Fig. 8D); vertex with hind margin slightly carinate and obsolete median carina on disc extending posterad beyond hind margin; sides of vertex strongly carinate, laminate above eye and extending anteriorly to apex of cephalic process; sides of vertex slightly bisinuate in lateral view (Fig. 8C–D); frons with 2 carinae extending ventrally on sides of process (Fig. 8E); cephalic process about
3.2 times as long as diameter of eye, projecting anterodorsad, curved, elongate and narrow; apical half ventrally with median carina getting laminate near apex (Fig. 8D–E); transverse wrinkles at base of process (Fig. 8C); postclypeus with sides strongly carinate and slight median carina; anteclypeus with median carina (Fig. 8E); labium very long, reaching or surpassing apex of abdomen.

**Thorax.** Yellow-brown with slightly paler markings (Fig. 8C–E); pronotum with strong median carina (Fig. 8C); carina very slightly sinuate in lateral view (Fig. 8D) and with strongly impressed point on each side (Fig. 8C); hind margin elevated above level of mesonotum medially (Fig. 8D); peridiscal carina anteriorly (Fig. 8C); sides of prothorax with oblique carina; lateral lobe of pronotum rounded posteriorly (Fig. 8D–E); mesonotum with median carina stopped at scutellum; curved peridiscal carina; short oblique carina at base of scutellum; scutellum transversely wrinkled (Fig. 8C).

**Tegmina** (Fig. 8A). Brown with irregular black-brown, often confluent, small spots with pale yellow-brown center; center of spots covered with white waxy secretion in fresh specimens; no spots beyond nodal line of cross-veins except at apicosutural angle; apex of clavus with large black-brown marking followed by white patch along sutural margin; costal margin straight, slightly rounded after nodal line; apical margin oblique, straight in middle and with angles rounded; sutural margin sinuate.

**Hind wings** (Fig. 8A). Broad; disc largely bright red or orange; apex and sutural margin brown-black; 2–4 small, brown-black spots on disc; sutural margin bisinuate.

**Legs** (Fig. 8A). Elongate and slender; pro- and mesofemora brown with 3 narrow, sinuate and often incomplete pale yellow rings; pro- and mesotibiae brown with 2 pale yellow rings; metafemora brown with imprecise yellow ring; metatibiae and metatarsi yellow-brown, tibiae darker basally; metatibiae with 5 (sometimes 4) lateral and 7 apical spines; 8–9 spines apicoventrally on first metatarsomere.

**Abdomen** (Fig. 8A). Red dorsally, brown-black ventrally.

**Male genitalia**
Very finely granulose, dark brown, paler along sides of anal tube (Figs 26–27); pygofer higher than long and with posterior margin broadly rounded in lateral view (Fig. 26); anal tube slightly elongate, 1.31 times longer than broad at apex and with lateral margins sinuate in dorsal view (Fig. 27); posterior margin obliquely rounded and underside nearly straight in lateral view (Fig. 26); gonostyli elongate, 1.6 times longer than high, not surpassing anal tube and broadly rounded at apex in lateral view (Fig. 26); ventral margin slightly sinuate on basal ⅔ (Fig. 26); dorsal margin obliquely directed dorsally on basal half, then straight after angle, pointed hook-like tooth at half of basal oblique part curved lateroventrally (Fig. 26); gonostyli nearly not visible from above (Fig. 27).

**Remarks**
Male genitalia without basodorsal process on gonostyli.

_Belbina madagascarensis_ can be separated (1) from _B. bergrothi_ by the less elongate gonostyli, without large pale marking ventrally and more broadly rounded apex, and less elongate anal tube; (2) from _B. foliacea_ by having gonostyli more broadly rounded apically and without a strong angle above the mediiodorsal tooth; (3) from _B. nympha_ by having the margins of the anal tube not rounded laterally and the dorsal margin of the gonostyli nearly straight on the apical half; (4) from _B. recurva_ by having the anal tube not produced into a semi-circular lateral plate.

**Distribution**
Fig. 46.
**Belbina nympha** (Stål, 1866) comb. nov.  
Figs 9A–E, 28–29, 50

*Cornelia nympha* Stål, 1866: 142 (type in NHRS).  
*Enchophora atomaria* Brancsik, 1893: 253, pl. 11: figs 7, 7a (type in HNHM). syn. nov.

*Enchophora atomaria* – Metcalf 1947: 114 (catalogued).  
*Cornelia atomaria* – Lallemand 1959: 88, fig. 35 (key, description, lateral view of head). — Constant 2004b: 31 (listed).

**Diagnostic characters**

(1) disc of hind wings red or orange (Fig. 9A); (2) ground colour of tegmina brown (Fig. 9A); (3) small-sized (less than 22 mm long); (4) cephalic process directed anterodorsad (Fig. 9D); (5) abdomen with 2 rows of black spots (Fig. 9A).

LT: ♂ (n = 3) 18.9 mm (18.0–19.8); ♀ (n = 3) 20.6 mm (19.6–21.9).

**Material examined**

**Type material**  
MADAGASCAR: Holotype of *Cornelia nympha*, ♀, [Madag.] [Stål] [Cornelia Stål] [Typus] [NHRS-HEMI 000000106] (NHRS).  

**Additional material**  
MADAGASCAR: 1 ♂, Antongil Bay, Mocquerys, 15°45’ S, 49°50’ E (HNHM); 1 ♀, Diego-Suarez Prov., forest area 7 km N of Joffreville, 12°20’ S, 49°15’ E, 360 m, 22–26 Jan. 2001, Malaise trap, Irwin, Schlinger & Harin’Hala (CAS); 1 ♂, Nosy-Be (BMNH); 1 ♀, Diego-Suarez, coll. De Bergevin (MNHN); 1 ♂, no data (probably Montagne d’Ambre), Feb., coll. Sicard (MNHN); 1 ♀, Maroantsetra, 15°26’ S, 49°44’ E (FSAG); 1 ♂, Nosy Komba, flanc, May 1956, A.R., Institut Scientifique Madagascar, Nosy Komba (Nosy Koba), 12°12’ S, 49°16’ E (FSAG); 1 ♀, Nosibé (FSAG); 1 ♀, Madagascar (MNPC).

**Male genitalia**

Very finely granulose, dark brown, with gonostyli slightly paler dorsally and ventrally (Figs 29–30); pygofer higher than long and with posterior margin broadly sinuate in lateral view (Fig. 29); anal tube elongate, 1.43 times longer than broad at apex and with lateral margins diverging on basal half, then broadly rounded in dorsal view (Fig. 30); curved ventrally and with posterior margin acutely rounded apically in lateral view (Fig. 29); gonostyli elongate, 1.63 times longer than high, not surpassing anal tube and broadly rounded at apex in lateral view (Fig. 29); all margins broadly rounded except dorsal margin obliquely straight on basal half (Fig. 29); lateral hook-shaped tooth at base of dorsal margin curved lateroventrally (Fig. 29); gonostyli nearly not visible from above (Fig. 30).

**Remarks**

Male genitalia without basodorsal process on gonostyli.
Belbina nympha can be separated (1) from B. bergrothi by the less elongate gonostyli, without large pale marking ventrally and more broadly rounded apex, and the less elongate anal tube; (2) from B. foliacea by having gonostyli more broadly rounded apically and without a strong angle above the mediadorsal tooth; (3) from B. madagascariensis by having the margins of the anal tube rounded laterally and the dorsal margin of the gonostyli broadly rounded on the apical half; (4) from B. recurva by not having the anal tube produced into a semi-circular lateral plate.

Distribution
See Fig. 50.

Belbina pionneaui Lallemand, 1922
Figs 10A–E, 30–31, 51

Belbina pionneaui Lallemand, 1922: 63 (type in FSAG).


Diagnostic characters
(1) disc of hind wings red (Fig. 10A); (2) tegmina dark red with membrane pale brown, numerous black-brown spots (Fig. 10A); (3) cephalic process broad, strongly curved dorsad (Fig. 10D–E); (4) large-sized (more than 25 mm long).

LT: ♂ (n = 3) 25.6 mm (25.1–26.0); ♀ (n = 13) 30.6 mm (25.8–32.9).

Material examined

Holotype
MADAGASCAR: ♂, [Type] [Manjakandriana, Madagascar] [Belbina pionneaui Lallem.] (FSAG).

Paratypes
MADAGASCAR: 2 ♂♂, 6 ♂♀, [Manjakandriana, Madagascar] [Para-type] (FSAG); 1 ♂, [Manjakandriana, Madagascar] [Para-type] [Belbina pionneaui Lall, V. Lallemand det., 1959] (NHRS); 1 ♂, [Manjakandriana, Madagascar] [Para-type] [Belbina pionneaui Lallemand, V. Lallemand det., 1956], 18°55' S, 47°48' E (NHRS).

Note: Lallemand (1922, 1959) stated that the type and paratypes are in his collection. It seems, however, that he subsequently gave one of the paratypes to NHRS.

Additional material
MADAGASCAR: 1 ♂, Madagascar (RBINS); 2 ♂♀, Tananarive, Lamberton, Tananarive (Antananarivo), 18°55' S, 47°31' E (RBINS); 1 ♂, 1 ♀, [“Ambatol...” (unreadable)], Lamberton (RBINS); 1 ♂, Maroantsetra, Lamberton, 15°26’, S, 49°44’ E (RBINS); 1 ♂, 4 ♀♀, Tananarive (NCSU, 1 ♀*: MNHN); 2 ♂♂, 1 ♀, no data (NCSU). – The specimen marked with * bears a label [Belbina pionneaui, cotype, Lallemand]. This specimen is not recognized as a paratype, as Lallemand (1959) stated that all type specimens were from Manjakandriana.

1 ♂ mislabeled “Soekaboemi, Java” has also been examined (INHS).

Examined on photographs
MADAGASCAR: 4 specimens, Tananarive, Lamberton (RMNH); 2 specimens, Maroantsetra, Lamberton (RMNH).
Male genitalia
Dark red (Figs 30–31); pygofer higher than long and with posterior margin straight, slightly rounded on dorsal ¼ in lateral view (Fig. 30); anal tube slightly elongate, 1.28 times longer than broad at apex and with lateral margins bisinuate in dorsal view (Fig. 31); posterior margin broadly rounded in lateral view (Fig. 30) with hind margin of anal opening slightly projecting posteriorly (Fig. 30) and hiding lateral margin apically in dorsal view (Fig. 31); gonostyli elongate, 1.35 times longer than high (dorsal process included), strongly surpassing anal tube and rounded at apex in lateral view (Fig. 30); ventral margin straight on basal ⅓, then sinuate (Fig. 30); dorsal margin with basal digitiform process slightly curved posteriorly and projecting laterodorsally, strong hook laterally at middle of process projecting ventrally (Figs 30–31); digitiform process narrowing from base to apex and acutely rounded apically (Figs 30–31).

Remarks
_Belbina pionneaui_ is a member of the _falleni_ group, showing a strong basodorsal digitiform process on the gonostyli. It can be separated from all species of the group by having the basodorsal digitiform process of the gonostyli narrowing from the base to the apex and the apical margin of the anal tube not notched under the anal opening in lateral view.

Distribution
See Fig. 51.

**Belbina recurva** Lallemand, 1950
Figs 11A–E, 32–33, 40–42, 52

_Belbina recurva_ Lallemand, 1950: 84 (type in FSAG).

_Belbina recurva_ – Lallemand 1959: 90, fig. 37a–c (key, description, lateral view of head, male genitalia). — Constant 2004b: 31, fig. 3 (listed, habitus).

Diagnostic characters
(1) disc of hind wings red (Fig. 11A); (2) ground colour of tegmina variegated brown (Fig. 11A); (3) veins of tegmina elevated (Fig. 11A); (4) large-sized (more than 26 mm long); (5) cephalic process very strongly curved and projecting posterodorsad (Fig. 11D–E); (6) abdomen with 2 rows of black spots dorsally (Fig. 11A).

LT: ♂ (n = 4) 27.3 mm (25.6–28.4); ♀ (n = 2) 29.8 mm (29.8–29.9).

Material examined
Holotype
MADAGASCAR: ♂, [Maroantsetra, Madagascar, Lamberton] [Type] [Belbina recurva Lallem., V. Lallemand det.]; 15°26’ S, 49°44’ E (FSAG).

Paratype
MADAGASCAR: 1 ♂, [Maroantsetra, Madagascar, Lamberton] [Paratype] (FSAG).

Additional material
MADAGASCAR: 1 ♀, E Madagascar, Sambava district, Marojejy, Ambinanitelo, 500 m, Dec. 1958, Raharizonia, 15°21’ S, 49°35’ E (RBINS); 1 ♂, Fianarantsoa, Ranomafana National Park, Talatakely, trail FF, 915–1000 m, 4–20 Nov. 1998, Lee & Ribardo, 21°15’ S, 47°27’ E (CAS); 1 ♀, E Madagascar,
Ambodivoangy, Jan. 1960, J. Vadon, 15°17'50" S, 49°36'47" E (MRAC); Antsianaka forest, 1964, 17°30' S, 48°30' E (MRAC); 1 ♂, 1 ♀, Madagascar, E coast, Grandidi, 1309-71 (MNHN); 1 ♂, E Madagascar, Mananara district, N Antanambe, Vadon & Peyrieras, 16°26' S, 49°51' E (MNHN); 1 ♂, E Madagascar, massif du Marojejy (rés. nat. intégr. 12), Ambatomitatao, 400 m, Jan. 1973, A. Peyrieras, 13°47'30" S, 49°51' E (MNHN); 1 ♀, Antongil Bay, Mocquerys, 15°45' S, 49°50' E (HNHM); 1 ♀, Antsiranana, Parc National de Marojejy, Manantenina River, 28.0 km 38° NE of Andapa, 8.2 km 333° NNW of Manantenina, 14°26'12" S, 49°46'30" E, 450 m, Malaise trap in rainforest, 12–25 Nov. 2003, B.L. Fischer et al. (CAS); 2 ♀♀, Rogez, 18°48' S, 49°42' E.

**Examined on photographs**

MADAGASCAR: 1 specimen, Masoala Peninsula, 30 May 2009, Dante Fenolio (Fig. 42), 15°30'48" S, 50°07'20" E; 1 specimen, Anajanaharibe Sud, 2 May 2010, Becky Rowe (Fig. 41), 14°46' S, 49°26' E; 1 specimen, Marojejy, near Cascade Humbert, 22 Nov. 2005 at 20:41, David C. Lees (Fig. 40), 14°26' S, 49°42' E.

**Male genitalia**

Black-brown with gonostyli slightly paler (Figs 32–33); pygofer higher than long and with posterior margin nearly straight in lateral view (Fig. 32); anal tube slightly transverse, 0.93 times as long as broad and with margins strongly projecting laterally on basal ⅔ into semi-circular lamina in dorsal view, apical ⅓ with lateral margins diverging towards apex (Fig. 33); apical margin acutely rounded in lateral view (Fig. 33); gonostyli elongate, 1.93 times longer than high, reaching apex of anal tube (Fig. 32); ventral margin broadly rounded to apex and dorsal margin strongly sinuate and with ante-apical notch in lateral view (Fig. 32); dorsal margin with strong, hook-shaped tooth at midpointing anteroventrally (Fig. 33); dorsal margin project laterally, semi-circular in dorsal view, with basal lobe and slightly folded internally near notch (Fig. 33).

**Remarks**

Male genitalia without basodorsal process on gonostyli.

*Belbina recurva* can be separated from all other species by having the anal tube produced into a semi-circular lateral plate.

**Distribution**

See Fig. 52.

*Belbina servillei* (Spinola, 1839)

Figs 12A–E, 34–35, 53

*Enchophora servillei* Spinola, 1839: 227, pl. 2, fig. 3a–c (neotype in HMNH).


*Phrictus servillei* – Schaum 1850: 65 (listed).


non *Belbina servillei* – Lallemand 1959: 88, fig. 36a–c (key, description, lateral view of head, male genitalia) – [Misidentification of *Belbina laetitiae* sp. nov.]
Diagnostic characters

(1) disc of hind wings turquoise (Fig. 12A); (2) head, pro- and mesonotum red (Fig. 12C–E); (3) ground colour of tegmina bluish-green (Fig. 12A); (4) cephalic process strongly curved, projecting dorsad (Fig. 12D).

LT: ♂ (n = 1) 20.4 mm; ♀ (n = 13) 23.3 mm (21.7–24.8).

Material examined

Neotype

MADAGASCAR: ♂, neotype of Belbina servillei (Spinola, 1839), here designated, [Madagascar, Antongil B., Mocquerys], 15°45′ S, 49°50′ E (HMNH).

A neotype is designated here in order to ensure stability in the nomenclature of the group, following rule 75.3 of the International Code of Zoological Nomenclature (ICZN 1999). The origin of the specimen described by Spinola (1839) is unknown and Stål (1866) was the first to give a location for the species (Madagascar). Lallemand (1959) designated a lectotype in his collection (FSAG) that is invalid according to nomenclatural rules. Furthermore, the only labelled specimen of this species, [Belbina servillei Sign., V. Lallemand det., 1963], in Lallemand’s collection is in fact a specimen of Belbina laetitiae sp. nov., as demonstrated by the genitalia illustrated in Lallemand (1959). In order to avoid further nomenclatural confusion, a neotype is designated which perfectly matches the illustrations given by Spinola (1839) and which is deposited in a public collection (HMNH). I chose a male with genitalia in good condition to facilitate further work on the species.

Additional material

MADAGASCAR: 1 ♂, 2 ♀♀, no data (MNHN); 1 ♀, near Tamatave, A. Raffray, 1884, Tamatave (Toamasina), 18°10′ S, 49°23′ E (MNHN); 1 ♂, 3 ♀♀, Madagascar, coll. De Bergevin (MNHN); 1 ♀, Tamina, coll. De Bergevin, 18°30′ S, 49°16′ E (MNHN); 3 ♀♀, E Madagascar, forest, coll. De Bergevin (MNHN); 1 ♀, Diego-Suarez, Ch. Alluauad, 1893, Diego Suarez (Ampanolahamirafy), 12°16′ S, 49°17′ E (MNHN); 2 ♀♀, Manjakandriana, 18°55′ S, 47°48′ E (FSAG); 1 ♂, 2 ♀♀, Manjakandriana, 16°24′ S, 49°51′ E (MNHN); 1 ♂, 2 ♀♀, Manjakandriana, 18°55′ S, 47°48′ E (FSAG); 2 ♂♂, 1 ♀, Madagascar (NMW, NHRS, ♀: NCSU); 2 ♂♂, Maroantsetra (NCSU).

1 ♂ mislabelled “S. America, Coll. Signoret” has also been examined (NMW).

Male genitalia

Brown (Figs 34–35); pygofer higher than long and with posterior margin concave in middle in lateral view (Fig. 34); anal tube as long as wide at apex and with lateral margins broadly produced in middle in dorsal view (Fig. 35); posterior margin of anal tube acutely rounded posteriorly in lateral view (Fig. 34) and hiding lateral margin apically in dorsal view (Fig. 35); gonostyli elongate, 1.33 times longer than high (including dorsal process, 1.88 times without process), strongly surpassing apex of anal tube and rounded at apex in lateral view (Fig. 34); ventral margin rounded on basal ½, then nearly straight (Fig. 34); dorsal margin with basal, strong, laminate process projecting dorsally, strong hook laterally at middle of process projecting ventrally; dorsal half of process slightly excavate and narrowly rounded dorsally, dorsal margin of gonostyli bisinuate after process (Fig. 34); gonostyli minutely rugulose on ventral half (Fig. 34).

Remarks

Belbina servillei can be separated (1) from all species of the B. falleni+ group by the brown colour of the genitalia, the laminate basodorsal digitiform process on the gonostyli and having the gonostyli
rounded at the apex, with the dorsal margin sinuate after the process; (2) from *B. bergrothi*, *B. foliacea*, *B. madagascariensis* and *B. nympha* by the basodorsal laminate process on the gonostyli; (3) from *B. bourgoini* sp. nov. by the laminate basodorsal process of the gonostyli.

**Distribution**

See Fig. 53.

**Discussion**

The genus *Belbina* contains most of the species of Fulgoridae recorded from Madagascar, 12 out of 17 (Constant 2004b), with the 5 other species belonging to the genera *Antsalovasia* Constant, 2004, *Radamana* Distant, 1906 and *Zanna* Kirkaldy, 1902. *Belbina*, *Antsalovasia* and *Radamana* are endemic to Madagascar while *Zanna* is also found in Afrotropical and Oriental regions.

Although the species of *Belbina* often have a wide distribution range (see maps on Figs 46–53), they are not regularly collected, probably due to their cryptic colour when sitting on tree trunks (see Figs 36, 38, 42). More field work should be conducted to document their life-history and to make observations on the host plants and the biology of these insects. Although they are actively searched by collectors, the feeding habits, larval stages and eggs remain unknown for all species.

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**References**


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